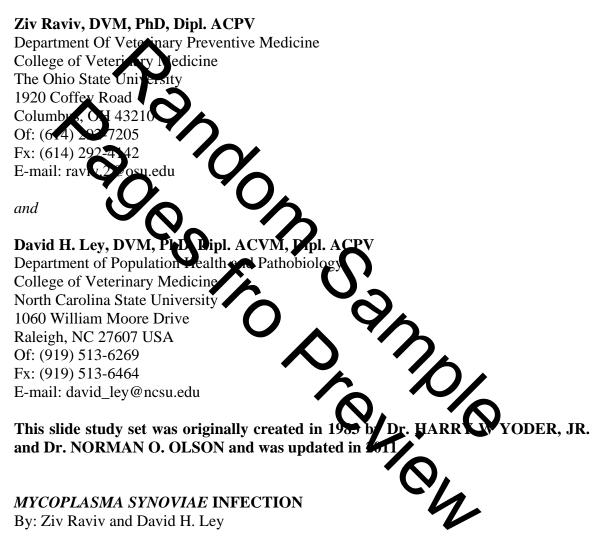
## MYCOPLASMA SYNOVIAE INFECTION

Slide study set #12

Prepared by:



*Mycoplasma synoviae* (MS) is a pathogen of chickens and turkeys, causing significant economic losses to poultry producers worldwide. Infection can be associated with upper respiratory disease, airsacculitis, synovitis, tenosynovitis, and bursitis. Disease severity has been influenced by other respiratory pathogens (e.g., Newcastle disease virus, infectious bronchitis virus), more virulent MS strains, and host species predilection (turkeys more susceptible than chickens).



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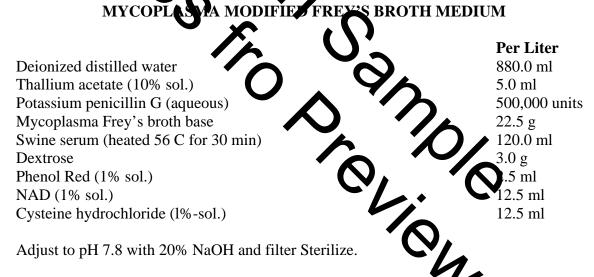
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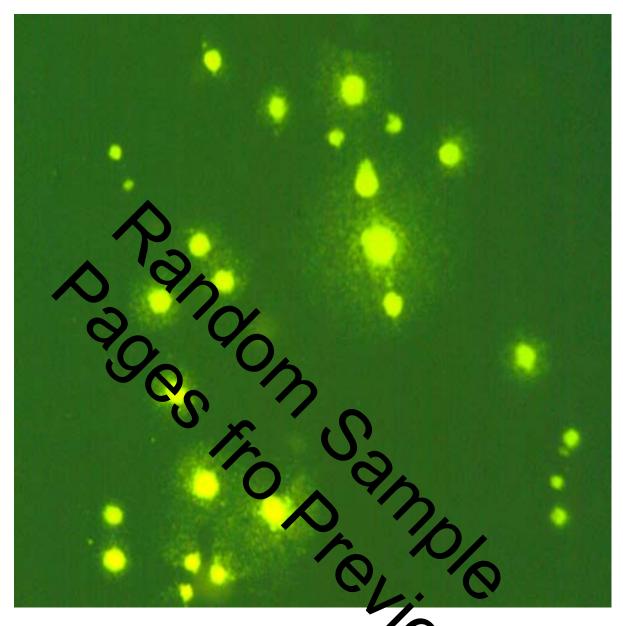
**History, Distribution, and Incidence**. Infectious synovitis was first described and associated with mycoplasma infection by Olson *et al.*, during the early 1950's. The causative organism was designated as Avian Mycoplasma Serotype S by Dierks et al., in 1967 and subsequently confirmed as a separate species, *Mycoplasma synoviae*, by Jordan *et al.*, in 1982. Soon after its identification MS appeared to have worldwide distribution. During the 1950's and 1960's the synovitis form of the disease was observed primarily in growing meat-type chickens (broilers), while since the 1970's the respiratory form of the disease has been seen more frequently. Flocks of laying chickens are commonly infected with MS with mild or subclinical signs. The disease usually appears in turkey flocks 10 to 20 weeks of age, primarily in multiage farms and in endemically MS infected areas. The outcome of infection is significantly affected by management factors, other respiratory pathogens (e.g., wwe stel disease virus, infectious bronchitis virus), virulence of the involved MS strains and the host species (turkeys more susceptible than chickens).

checause of infectious synovitis of chickens and turkeys. MS Etiology is the d more freque ersistent infection of the upper respiratory tract which itly produces a plved with air accuritis. MS is a very fastidious cell wall-less bacterium sometimes i 10-15% swine serum, and specifically requires the requiring a ch mediu ide adenine diny eleotide (NAD). Updated specifications for MS addition of nic otina culture follow



Add thallium acetate to the water first to avoid precipitation of koteins of media and serum. Horse serum is adequate for MG, but swine serum is best for MS. Cysteine hydrochloride is added to reduce the NAD (beta nicotinamide adenine dinucleotide) which is required for the growth of MS. For agar plates 1.5% agar is used. For potentially contaminated specimens, an extra 20 ml of 1% thallium acetate and 2,000,000 units of penicillin per liter may be added. Ampicillin (from 200-1000mg/l) may be substituted for penicillin.

**Colony morphology**. Colonies on solid media are best observed with a dissecting microscope at 30X magnification using indirect lighting. They appear as raised, round,



**SLIDE 4**. Mycoplasma cultures isolated from trachea swals or air-sac lesions must be speciated. *M. synoviae* and *M. gallisepticum* are only 2 of over 20 possible species of avian mycoplasma. This slide shows the greenish glow of colonies that are positive in the immunofluorescence (IF) test using species-specific antibodies (slide courtesy of Dr. Stanley H. Kleven).



**SLIDE 5**. Chicken with its tongue pulled aside to position the larynx for insertion of a swab to obtain tracheal exudate for cultivation of most avian mycoplasma, including MS. (Slide courtesy of Dr. Stanley H. Kleven).



**SLIDE 6.** Normal air-sac membranes are so thin aid dear that they are almost invisible. The air sacs are primarily paired extensions (the fact, abdominal, etc.) of the air passages from the bronchioles on out beyond the lungs into various body cavity spaces. Some are within hollow bones. This slide shows a moderately inflamed air sac as noted by slight thickening of the membrane, some abouty evidate, and increasing flecks of yellowish caseous exudate as the process continues.

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